# SOUTH AUSTRALIA

## **AUStralian RIVer Assessment System**



## **ACCREDITATION MANUAL**













Department of the Environment and Hevitage

#### SOUTH AUSTRALIA AUStralian RIVer Assessment System (AUSRIVAS) **ACCREDITATION MANUAL**

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#### Front Cover Photograph:

Artemore Creek, Nildottie Spring in the Flinders Ranges. Photo by Chris Madden

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## **INTRODUCTION**

AUSRIVAS sampling methods are standardized within each State and Territory, but vary nationally. Therefore, AUSRIVAS Training and Accreditation Courses have been tailored specifically for each State and Territory. Additional training and accreditation may be required to use AUSRIVAS as an accredited operator outside the state or territory where accreditation was originally obtained.

The South Australian AUSRIVAS Training and Accreditation Course has been developed to provide uniformity and consistency in the application of AUSRIVAS methods in South Australia. The course comprises two major components, a training component and an accreditation component. Each of these components consists of four modules (listed below). Depending on the needs of the course participant, one or all of the modules may be completed (i.e., training with or without accreditation or just accreditation).

#### Module 1

#### **Pre-Field & Site Information**

- Preparation for field work
- Collection of site information

#### Module 2

#### **Field Work**

- Collection of field data
- Macroinvertebrate sampling

#### Module 3

#### Laboratory

- Sample processing laboratory sub-sample
- Macroinvertebrate Identification
- Data entry (Training Component only)
- Quality Assurance/Quality Control (Training Component only)

#### Module 4

#### **AUSRIVAS Predictive Models**

- Using the predictive models
- Interpreting the results

The South Australian AUSRIVAS Accreditation Manual provides details of the accreditation component, including the assessment sheets and instructors notes for each of the four modules. The manual also provides a guide to the level of training required to become accredited. For more information on the training component, see the AUSRIVAS Training and Accreditation Course Outline, Content and Structure document, or visit the AUSRIVAS web site at <a href="http://ausrivas.canberra.edu.au">http://ausrivas.canberra.edu.au</a>.

#### **Assessment Sheets**

Assessment sheets have been provided for each of the four modules. Each assessment sheet details a number of exercises that course participants must pass to be accredited as a South Australian AUSRIVAS operator. Assessment sheets should be photocopied from the manual or printed from the South Australian AUSRIVAS Training and Accreditation CD and handed out to course participants at the beginning of each accreditation module.

#### **Instructor's Notes**

For each accreditation module, instructor's notes have been provided to aid instructors in preparing, conducting and assessing the accreditation exercises. The notes also ensure a standardized accreditation procedure among instructors. Included in the instructor's notes are Mark Allocation Sheets. These sheets provide the instructor with the marking scheme to be used when assessing exercises.

#### Accreditation Assessment Results Summary Sheet

An Accreditation Assessment Results Summary Sheet (Appendix 1) is included in the South Australian AUSRIVAS Accreditation Manual to summarize the accreditation assessment results for each course participant.

## **MODULE 1 – ASSESSMENT SHEET**

#### Page 1 of 1

Name: \_\_\_\_\_

**Pre-Field & Site Information** 

Collection of Site Information

**Exercise 1. Site Information** 

Complete the "Site Description" and "Catchment Variables" sections of a field sampling sheet for sites \_\_\_\_\_\_, \_\_\_\_, and \_\_\_\_\_.

(Exercise 1 Mark = \_\_\_%).

Note: A mark of 90% or greater (averaged over the three sites) is required in Exercise 1 to pass Module 1.

## **MODULE 1 - INSTRUCTORS NOTES**

#### Pre-Field & Site Information

A list of materials and resources required for Module 1 is given below.

- South Australian Sampling and Processing Manual
- Field Sampling Sheets
- Maps (laminated and non-laminated)
- Map measuring devices eg. digital planimeter, map wheel, ruler etc.
- Magnifying glass
- GPS (for converting grid references to latitude and longitude)

#### Collection of Site Information

#### **Exercise 1. Site Information**

#### <u>Preparation</u>

Before conducting Exercise 1, the instructor must complete the "Site Description" and "Catchment Variables" sections of a field sampling sheet for each of the three sites being assessed. The information collected by the instructor will become the answers against which the course participant's site information will be assessed. The sites used in the Module 1 (and Module 2) assessment should be reference sites from the area in which the course is being conducted. The use of local sites increases the information retained by the participants because the information is more relevant. The use of the same sites throughout the accreditation process provides continuity between accreditation modules, and enables course participants to gain a detailed knowledge of the sites being examined.

Copy the site numbers of the three sites being assessed onto the Module 1 assessment sheet. For each site, place a dot on the relevant map indicating the site's location.

#### <u>Assessment</u>

Attach copies of the AUSRIVAS accreditation Module 1 assessment sheet and mark allocation sheet No. 1 to three blank field sampling sheets and hand out to each course participant. Participants must complete the "Site Description" and "Catchment Variables" sections of a field sampling sheet for each site. Once participants have completed the exercise, collect sheets, ensuring participant's names are on the assessment sheet.

#### <u>Marking</u>

Award marks for each question according to the marking scheme on mark allocation sheet No. 1. An answer is marked as correct if it is the same as the instructor's answer, or for variables listed in Table 1, within the acceptable error range. Answers for variables not listed in Table 1 must be the same as the instructor's answer to be marked correct (i.e., no variation is allowed). Answers receive full marks if correct, and zero marks if incorrect (i.e., a proportion of a question's mark cannot be given). Missing values are to be marked as wrong even when the correct answer is zero.

## Mark Allocation Sheet No. 1

## **GRAND TOTAL = 67**

#### MRHI Field Data Sheet

SITE DESCRIPTIC	ОN	5	Location	Name (river	and site lo	cation)
Location Code		5	_			
Mapsheet Name		1	_Mapsheet	no and sca	le	
AMG Zone 3	Easting		3	Northing		3
Latitude(	5	Longtitud	e	5	Altitude	5 m
Nearest Named Place	e (1)					

#### Catchment Variables

.

River 5	Location Code	5		
Stream Order 5	(1:50,000 scale map)			
Distance From Source (DFS)	5km			
Catchment Area Upstream	5 km <sup>2</sup>			
Slopem/last km (1:50,000 scale map)				

Variable	Acceptable Error
Easting	±100m
Northing	±100m
Latitude	$\pm 00^{\circ} 00' 04''$
Longitude	±000° 00' 04"
Altitude	±20m
Distance from Source	$\pm 10\%$
Catchment Area Upstream	$\pm 10\%$
Slope	± 10%

**Table 1.** The acceptable error for specific variables in the "Site Description" and "Catchment Variables" sections of a field sampling sheet. Variables not listed below must be the same as the instructor's answer to be marked correct.

A mark of 90% or greater (averaged over the three sites) is required in Exercise 1 to pass Module 1. Record the mark for Exercise 1 in the space provided on the assessment sheet, and on the participant's Accreditation Assessment Results Summary Sheet (Appendix 1). Hand the assessment sheet and field sampling sheets back to participants and work through any areas of difficulty with them. Participants should retain their field sampling sheets for use in Module 2.

## **MODULE 2 – ASSESSMENT SHEET**

#### Page 1 of 1

Name:

Field Work

Collection of Field Data

#### **Exercise 1. Field Sampling Sheet**

Complete a field sampling sheet for sites \_\_\_\_\_\_ and \_\_\_\_\_. A mark of 90% or greater (averaged over the two sites) is required to pass Exercise 1.

(Exercise 1 Mark = \_\_\_%).

#### Macroinvertebrate Sampling

#### **Exercise 2. Sample Collection**

Collect a 10m riffle and edge sample from site \_\_\_\_\_\_ and place into separate labelled sample containers. A mark of 90% or greater (averaged over the two habitats) is required to pass Exercise 2.

As	sessable Criteria	Riffle Mark	Edge Mark
٠	10m of habitat sampled	/ 2	/ 2
٠	Available range of micro-habitats sampled	/ 5	/ 5
٠	Habitat sampled effectively	/ 3	/ 3
٠	Net rinsed when necessary	/ 2	/ 2
٠	Sample transferred successfully from net to sample container	/ 4	/ 4
٠	Sample container correctly labelled	/ 2	/ 2
•	Sample preserved correctly with operator wearing protective gloves, goggles and clothing (as required).	/ 2	/ 2
	Total	/ 20	/ 20

(Exercise 2 Mark = %).

Note: A mark of 90% or greater is required in both exercises to pass Module 2.

## **MODULE 2 - INSTRUCTORS NOTES**

## **Field Work**

A list of materials and resources required for Module 2 is given below.

ITEM	QUANTITY	CHECK
South Australian Sampling and Processing Manual		
Triangular Kicknets, inner frame dimension 300mm with a 250 um mesh bag		
Sieves, 250 um mesh for "rapid" sampling		
Small plastic bucket		
Medium white tray		
Formaldehyde - buffered & with Rose Bengal stain		
Gloves		
Bug sampling containers		
250ml plastic bottles for water quality samples		
Flow meter		
Float ( for shallow streams where meter cannot be used)		
Waders + spare		
Wader repair kit		
Field meters for DO, pH, EC and Temp.		
Alkalinity kit		
Spare batteries		
Spare DO membranes & O <sub>2</sub> probe solution		
Calibration standards		
Calibration instructions		
GPS		
Digital camera		
Mobile phone		
Field Sampling Sheets		
Clipboard		
Maps		
Marker pens/ pencils		
Esky (large) & ice and/or fridge		
First Aid Kit		
4WD recovery equipment		
Tape measure 100m		
Camera & film		
Sunscreen		
Toilet paper / shovel		
Rain Jackets		
Plastic bags for veg. samples		
Labels and rubber bands		
Reference - "A Field Guide to Waterplants in Australia" Sainty and Jacobs 1994		

#### Collection of Field Data

#### **Exercise 1. Field Sampling Sheet**

#### <u>Preparation</u>

Before conducting Exercise 1, the instructor must complete field sampling sheets for the two sites being assessed. These sheets will provide the answers against which the course participant's field sampling sheets will be assessed. The sites should be selected from the three reference sites used in the Module 1 assessment.

Copy the site numbers of the sites being assessed onto the Module 2 assessment sheet.

#### <u>Assessment</u>

Attach copies of the AUSRIVAS accreditation Module 2 assessment sheet to mark allocation sheet No. 2 and hand out to each course participant. Participants should already have field sampling sheets from Module 1. The participants must complete a field sampling sheet for each site. Once participants have completed the exercise, collect the field sampling sheets, ensuring participant's names are on the front page. Participants should retain their Module 2 assessment sheet until all Module 2 exercises have been completed.

#### <u>Marking</u>

Award marks for each question according to the marking scheme on mark allocation sheet No. 2. An answer is marked as correct if it is the same as the instructor's answer, or for variables listed in Table 2, within the acceptable error range. Answers for variables not listed in Table 2 must be the same as the instructor's answer (or within the same category) to be marked correct. Answers receive full marks if correct, and zero marks if incorrect (i.e., a proportion of a question's mark cannot be given). Missing values are to be marked as wrong even when the correct answer is zero.



#### Chemical and Physical Description



#### Stream Transects

2 m 3 m 7 m 1 6 m 9 m 10m 11m 12m 13m 14m Width 1 m 4 m 5 m 8 m Depth (cm) 3 15m 16m 17m 18m 19m 20m 21m 22m 23m 24m 25m 26m 27m 28m Depth (cm) 2 1 m 2 m 3 m 4 m 5 m 6 m 7 m 8 m 9 m 10m 11m 12m 13m 14m (3) Depth (cm) 18m 19m 20m 21m 22m 23m 24m 25m 26m 27m 28m 15m 16m 17m Depth (cm) 3 1 m 2 m 12m 13m 14 m 3 m 4 m 5 m 6 m 7 m 8 m 9 m 10m 11m Depth (cm) (3) 15m 16m 17m 18m 19m 20m 21m 22m 23m 24m 25m 26m 27m 28m Depth (cm)

#### **Habitat Descriptions**



#### **Biological Description**

Macrophytes 3 SUBMERGED/ FLOATING EMERGENT (ctd.) Carex (Tussock Sedge)	Dominant Terrestrial Vegetation		3			
SUBMERGED/ FLOATING       EMERGENT (cid.)         Azolla       Carex (Tussock Sedge)         Chara (Stonewort)       Crassula (Crassula)         Myriophyllum (Water Milfoll)       Cyperus (Sedge)         Nitela (Stonewort)       Eleocharis (Spikerush)         Potamogeton (Pondweed)       Rorrippe (Watercress)         Ruppia (Sea Tassel)       Polygonum (Smartweed)         Valisneria (Ribbonweed)       Solepis (Clubrush)         Zainchellis       Mimulus         Other       Bolboschoenus (Clubrush)         Zainchellis       Mimulus         Other       Schoenopictus (Clubrush)         Schoenopictus (Clubrush)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Other         Phragmites (Common Reed)       Juncus (Rush)         Juncus (Rush)       Galitriche (Starwort)         Calitariche (Starwort)       (1)         Algae (Attached/ Floating)       Cladophora       Spirogyra         Sample collected?       algae       macrophyte       (1)         Percent of 100 m reach covered by:       Periphyton       1<10% 210-35% 335-65% 4 65-90% 5 >90% (3)       (3)         Moss       1       10-35% 335-65% 4 65-90% 5 >90% (3)       (3)       (3)	Macrophytes 3					
Azolla       Carex (Tussock Sedge)         Chara (Stonewort)       Crassula (Crassula)         Myriophyllum (Water Milfoli)       Cyperus (Sedge)         Nitela (Stonewort)       Elecoharis (Spikerush)         Potamogeton (Pondweed)       Rorrippa (Watercress)         Potamogeton (Robonweed)       Rorrippa (Watercress)         Triglochin (Water Ribbon)       Ranuculus (Buttercup)         Valisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboschoenus (Clubrush)         Zanichellia       Mimulus         Other       Schoenopfectus (Clubrush)         Calistriche (Starwort)       Other         Phragmites (Common Reed)       Joncus (Rush)         Calitriche (Starwort)       (Idal 100%) (1)         Algae (Attached/ Floating)       Cladophora       Spirogyra         Sample collected?       algae       macrophyte       none (1)         Percent of 100 m reach covered by:       Periphyton       1       1         Periphyton       1       1       3       5-65% 4 65-90% 5 >90% (3)         Moss       1       1       2       10-35% 3 35-65% 4 65-90% 5 >90% (3)         Periphyton       1<10% 2	SUBMERGED/ FLOATING		EMERGEN	NT (ctd.)		
Chara (Stonewort)	Azolla		Carex (Tu	ssock Sedge	e)	
Myriophyllum (Water Milfoli)       Cyperus (Sedge)         Nitelia (Stonewort)       Elecoharis (Spikerush)         Potamogeton (Pondweed)       Rorrippa (Watercess)         Ruppia (Sea Tassel)       Polygonum (Smartweed)         Yallisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboscheenus (Clubrush)         Zanichellia       Clubrush)         Zanichellia       Clubrush)         Cotula (Water Rubon)       Scheenoplectus (Clubrush)         California       Scheenoplectus (Clubrush)         Cotula (Waterbutton)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Phragmites (Common Reed)         Juncus (Rush)       Typha (Custhy)         Galitriche (Starwort)       Total 100%) (1         Algae (Attached/ Floating)       Cladophora       Spirogyra         Algae (Attached/ Floating)       Cladophora       Spirogyra         Sample collected?       algae       macrophyte       0         Moss       1       1       3       5-55% 4 65-90% 5 >90% (3)         Moss       1       1       1       1         Periphyton       1       1       1       1 </td <td>Chara (Stonewort)</td> <td></td> <td>. Crassula</td> <td>(Crassula)</td> <td></td> <td></td>	Chara (Stonewort)		. Crassula	(Crassula)		
Nitella (Stonewort)       Eleocharis (Spikerush)         Potagogeton (Pondweed)       Rorrippa (Watercess)         Ruppia (Sea Tassel)       Polygonum (Smartweed)         Triglochin (Water Ribbon)       Rannoulus (Buttercup)         Vallisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboschoenus (Clubrush)         Zanichellia       Mimulus         Other       Schoenoplectus (Clubrush)         Cotula (Waterbutton)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Tother         Juncus (Rush)       Galitriche (Starwort)         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Native vegetation       3         % Native vegetation       1         Algae (Attached/ Floating)       Cladophora         Spirogyra       Enteromorpha         sample collected?       algae         Moss       1         Filamentous algae       1<10%	Myriophyllum (Water Milfoil)		Cyperus (	Sedge)		
Potamogeton (Pondweed)       Rorrippa (Watercress)         Ruppia (Sea Tassel)       Polygonum (Smartweed)         Triglochin (Water Ribbon)       Ranunculus (Buttercup)         Zanichellia       Mimulus         Zanichellia       Mimulus         Other       Boiboschoenus (Clubrush)         Zanichellia       Mimulus         Other       Schoenoplectus (Clubrush)         EMERGENT       Other         Typha (Cumbungi)       Other         Pragmites (Common Reed)       Juncus (Rush)         Callitriche (Starwort)       Gatophora         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Native vegetation       3         3       (total 100%)       1         Algae (Attached/ Floating)       Cladophora       Spirogyra         Periphyton       1<10%	Nitella (Stonewort)		Eleocharis	(Spikerush	)	
Ruppia (Sea Tassel)       Polygonum (Smartweed)         Triglochin (Water Ribbon)       Ranunculus (Buttercup)         Vallisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboschoenus (Clubrush)	Potamogeton (Pondweed)		Rorrippa	(Watercress)		
Triglochin (Water Ribbon)       Ranunculus (Buttercup)         Vallisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboschoenoplectus (Clubrush)         Schoenoplectus (Clubrush)       Clubrush)         Cotula (Waterbutton)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Phragmites (Common Reed)         Juncus (Rush)	<i>Ruppia</i> (Sea Tassel)		Polygonur	<i>n</i> (Smartwee	ed)	
Valisneria (Ribbonweed)       Isolepis (Clubrush)         Zanichellia       Mimulus         Other       Bolboschoenus (Clubrush)         Schoenoplectus (Clubrush)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Other         Phragmites (Common Reed)       Juncus (Rush)         Juncus (Rush)       Galitriche (Starwort)         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Exotic vegetation       3         (total 100%)       1         Algae (Attached/ Floating)       Cladophora         Spirogyra       Enteromorpha         sample collected?       algae         Moss       1         Filamentous algae       1         Natrophytes       1         1       2         Moss       1         Flauna Observed       N/A	Triglochin (Water Ribbon)		. Ranunculı	<i>ıs</i> (Buttercu	p)	
Zanichellia       Mimulus         Other       Bolboschoenus (Clubrush)         Schoenoplectus (Clubrush)       Cotula (Waterbutton)         EMERGENT       Other         Typha (Cumbungi)       Phragmites (Common Reed)         Juncus (Rush)       Gallitriche (Starwort)         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Native vegetation       3         % Native vegetation       3         % Native vegetation       1         Algae (Attached/ Floating)       Cladophora         Springyra       Enteromorpha         sample collected?       algae         Moss       1         Filamentous algae       1         Nacrophytes       1         Status 2       10-35%         % Baserved       3         Status 2       10-35%         % Baserved       3         Status 2       10-35%         % Baserved       3         % Status 2       10-35%         % Status 2       10-35%         % Status 2       10-35%         % Status 2       10-35%	Vallisneria (Ribbonweed)		.Isolepis (C	Clubrush)		
Other       Bolboschoenus (Clubrush)	Zanichellia		Mimulus			
Schoenoplectus (Clubrush)         Cotula (Waterbutton)         EMERGENT         Typha (Cumbungi)         Phragmites (Common Reed)         Juncus (Rush)         Callitriche (Starwort)         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Native vegetation       1         % Native vegetation	Other		.Bolboscho	enus (Club	rush)	
Cotula (Waterbutton)         EMERGENT         Typha (Cumbungi)         Phragmites (Common Reed)         Juncus (Rush)         Calilitriche (Starwort)         Epiphyte Cover       Nil         Slight       Moderate         Extensive       1         % Native vegetation       3         % Exotic vegetation       3         (total 100%)       1         Algae (Attached/ Floating)       Cladophora         Spirogyra       Enteromorpha         sample collected?       algae         Moss       1         % Nass       1<10%			.Schoenop	<i>lectus</i> (Club	orush)	
EMERGENT       Other         Typha (Cumbungi)       Phragmites (Common Reed)         Juncus (Rush)       Callitriche (Starwort)         Callitriche (Starwort)       Slight         Moderate       Extensive         % Native vegetation       3         % Exotic vegetation       3         (1041 100%)       1         Algae (Attached/ Floating)       Cladophora         Sample collected?       algae         macrophyte       none         %       1         %       1         %       2         %       2         %       2         %       3         %       2         %       2         %       2         %       2 <t< td=""><td></td><td></td><td>.Cotula (W</td><td>aterbutton)</td><td></td><td></td></t<>			.Cotula (W	aterbutton)		
Typha (Cumbungi)       Phragmites (Common Reed)         Juncus (Rush)	EMERGENT		Other			
Phragmites (Common Reed)	Typha (Cumbungi)					
Juncus (Rush)       Callitriche (Starwort)         Callitriche (Starwort)       Slight       Moderate       Extensive       1         Epiphyte Cover       Nil       Slight       Moderate       Extensive       1         % Native vegetation       3       % Exotic vegetation       3       (total 100%)       1         Algae (Attached/ Floating)       Cladophora       Spirogyra       Enteromorpha         sample collected?       algae       macrophyte       none       1         Percent of 100 m reach covered by:       Periphyton       1       1         Moss       1       2       10-35%       3       35-65%       4       65-90%       5 >90%       3         Filamentous algae       1       10%       2       10-35%       3       35-65%       4       65-90%       5 >90%       3       3         Fauna Observed       N/A	Phragmites (Common Reed)					
Callitriche (Starwort)         Epiphyte Cover       Nil       Slight       Moderate       Extensive       1         % Native vegetation       3       % Exotic vegetation       3       (total 100%)       1         Algae (Attached/ Floating)       Cladophora       Spirogyra       Enteromorpha         sample collected?       algae       macrophyte       none       1         Percent of 100 m reach covered by:       Periphyton       1       1       3       3         Moss       1       10%       2       10-35%       3       35-65%       4       65-90%       5       >90%       3       3         Filamentous algae       1       10%       2       10-35%       3       35-65%       4       65-90%       5       >90%       3       3       3         Fauna Observed       N/A	Juncus (Rush)					
Epiphyte Cover       Nil       Slight       Moderate       Extensive       1         % Native vegetation       3       % Exotic vegetation       3       (total 100%)       1         Algae (Attached/ Floating)       Cladophora       Spirogyra       Enteromorpha         sample collected?       algae       macrophyte       none       1         Percent of 100 m reach covered by:       Periphyton       1<10%	Callitriche (Starwort)					
% Native vegetation       3       (total 100%)       1         Algae (Attached/Floating)       Cladophora       Spirogyra       Enteromorpha         sample collected?       algae       macrophyte       none       1         Percent of 100 m reach covered by:       Periphyton       1       1       3       3         Moss       1       10%       2       10-35%       3       35-65%       4       65-90%       5       90%       3       3         Moss       1       10%       2       10-35%       3       35-65%       4       65-90%       5       90%       3       3       3         Filamentous algae       1<10%	Epiphyte Cover Nil	Slight	Moderate		Extensive	
Algae (Attached/ Floating)       Cladophora       Spirogyra       Enteromorpha         sample collected?       algae       macrophyte       none       1         Percent of 100 m reach covered by:       Periphyton       1<10%	% Native vegetation	3	_% Exotic v	egetation	3	_(total 100%) 1
sample collected? algae macrophyte none 1 Percent of 100 m reach covered by: Periphyton 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Moss 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Filamentous algae 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Macrophytes 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Fauna Observed N/A	Algae (Attached/ Floating)	Cladophora	а	Spirogyra		Enteromorpha
Percent of 100 m reach covered by: Periphyton 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Moss 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Filamentous algae 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Macrophytes 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Fauna Observed N/A	sample collected? algae	macrophyt	е	none (1	)	
Periphyton       1<10%	Percent of 100 m reach covered	by:				
Moss 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Filamentous algae 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Macrophytes 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Fauna Observed N/A	Periphyton	1<10%	2 10-35%	3 35-65%	4 65-90%	5 > 90%
Filamentous algae 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% Macrophytes 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Fauna Observed	Moss	1<10%	2 10-35%	3 35-65%	4 65-90%	5 >90%
Macrophytes 1<10% 2 10-35% 3 35-65% 4 65-90% 5 >90% 3 Fauna Observed	Filamentous algae	1<10%	2 10-35%	3 35-65%	4 65-90%	5 >90%
Fauna Observed	Macrophytes	1<10%	2 10-35%	3 35-65%	4 65-90%	5 >90%
Fauna Observed						$\left( \begin{array}{c} 3 \end{array} \right)$
	Fauna Observed N	/A				

#### AUSRIVAS Accreditation - Module 2

General	Stream	Description	and	Use
---------	--------	-------------	-----	-----

			-				
WATER ODOURS:	normal	sewage	petroleum	chemical	none		
WATER OILS:	slick	sheen	globs	flecks	none	1	
TURBIDITY:	clear	slight	turbid	opaque	1		
PLUME (amount of fine s	ediment gene	rated when	sediment is	disturbed):	little	s o m e	lots 1
SEDIMENT ODOURS	normal	sewage	petroleum	chemical	anaerobic	none	
FLOW LEVEL: No flow	(relative to normal	inundation level, sl Low	hown by terrestrial	grasses, eroded ar Normal	ea or boundary in b	ank sediment) High	1
Are the undersides of s	stones which	n are not de	eply embec	lded black?		yes	no (1)
SEDIMENT DEPOSIT	S:	none	fine organic	s manure	sand	relict shells	1
EXISTENCE OF EROSI	O N :	none	s o m e	moderate	heavy	causes:	(
NPS POLLUTION:	no evidence	potential			obvious	type	(
RESTRICTIONS TO F	LOW	present types	upstream /	downstream		-	absent (
LANDUSE:	Native forest Residential	commercial	Forestry Industria	Native past	ure Recreational	Grazing other	Cropped
BARS: (bed surface pr	otruding fror	m normal wa	ater level &	forming a b	ar)	%	

#### Samples Collected

	No. Net Samples 1
1.25L water	Description of Net Samples
50mL Filtered sample	1]
Diatom > (1)	2
Protozoa [>5000 vS]	<u>3</u>
Photographs N/A Comments	N/A
Any Other Observations	)
	Page Total - 1

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]	HABIT	AT ASSI	ESSMENT	FIEI	LD DA	TA S	HEET		
Date:	5)	R	iver:		(5)				
Location Code:	(5)								
	(1)	•••••							
Name of recorder	<u>·</u>	••••••							
CHANNEL DESCR	<b>IPTION</b>								
Amount of stable habita	it >	50%	30-50%	10-30%	D	<10%			(3)
Description of main hab	itat (sedime	ent category a	nd/or macrophyt	e type)			•••••		3
Embeddedness (%grave	l, cobble, b	oulders surrou	inded by fine sed	liment)	N/A	0-25	25-50 50-75	>75	3
Velocity/depth categorie	s present shallow ri	deep (> ffle	30cm) edge	shallow	v edge	deep (>1	0cm) riffle		3
Channel width/depth ra	tio (high flo	w bank width	, top of bank to b	oottom of	water)				(3)
Bottom scouring	ABSENT	PRESE	NT (amount)						$\overbrace{3}$
Bonk slong/stoopness (re	$n_{0} of^{0}$	T REDE							
Dank Slope/Steepness (12	inge 01 )	••••••	••••••	•••••					$(\cdot)$
Bed Condition (may use	e more than	one category)	3						
amount of erosion or	deposition	location in b	ed		particle s	ize	% of reach	source	/cause
Nil (no erosion or depo	osition)								
Slight erosion									
Slight deposition									
Moderate erosion									
Moderate deposition									
Extreme erosion									
Extreme deposition									
Pools in reach BANK DESCRIPTI	Riffles in : <u>ON</u>	reach.	Adequacy of ha	abitat:		<u> </u>			
Bank stability									
stable (good veg cover, n limited erosion (good veg moderate erosion (discon extensive instability (littl extreme instability (evide	o significant cover, mine tinuous veg, e effective v nce of rapid	t damage to bar or isolated eros obvious dama eg, recent bank unchecked ero	nk structure, no ex ion, no continuou ge to bank structu movement, large ssion, little effecti	xposed roo is damage re, moder e amounts ve veg, at	ots) to bank, so ate exposu of exposed bundant exp	ome expo ire of roo d roots) posure of	osed roots) ts) Froots)	3	
Bank vegetative cover		>80%	50-79%	0	25-49%		<25%	3	
BUFFER DESCRIE	TION (ex	xclude grass	es)						
Width of buffer	<5m 5-	-10m	10-30m	30-40m	1 1	>40m	3		
<b>Longitudinal continuity</b> Percent of bank length wi Any gaps >10m long?	<b>of buffer</b> th >5m widt	h of vegetation	cover: left bank	3	)rig	ht bank	3		~
Buffer type and cover	>5m tall t	rees	continuous(>809	%)	patchy(2	0-80%)	sparse(<20%)	none	(3)
	<5m tall s	hrubs	continuous		patchy		sparse	none	3
	sedges, sa	mphire	continuous		patchy		sparse	none	$\overline{3}$
Evidence of regeneratio	n of indiger	nous species?	3				Page	Total = 7	79

AUSRIVAS Accreditation - Module 2

#### Field Data Sheet - Supplement

Catchment Variables



#### Chemical Description



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Variable	Acceptable Error
Depth Gauge (m)	± 10%
Conductivity ( $\mu$ S cm <sup>-1</sup> )	± 10%
Dissolved Oxygen (mg $l^{-1}$ )	± 10%
Dissolved Oxygen % sat.	± 10%
Water Temperature (°C)	± 10%
pH	± 10%
Secchi Depth (cm)	± 10%
Shade Cover %	± 10%
Stream width	± 20%
Area of Site – Riffle and Edge	± 10%
Current Speed – Max and Min	± 20%
Substratum description (% cover of	± 10%
each size category)	
% Algal Cover	$\pm 10\%$
% Detritus Cover	$\pm 10\%$
% Native Vegetation	± 10%
% Exotic Vegetation	± 10%
% Bars	± 10%
Channel width/depth ratio	± 10%
Bank slope/steepness	$\pm 10^{\circ}$
% erosion or deposition in reach	± 10%
Longitudinal continuity of buffer –	± 10%
left and right banks	
Alkalinity (mg $\Gamma^1$ )	± 10%
Carbonates ( $CO_2^{-2}$ mg $l^{-1}$ )	+ 10%

**Table 2.** The acceptable error for specific variables on the field sampling sheet. Variables not listed below must be the same as the instructor's answer (or within the same category) to be marked correct.

A mark of 90% or greater (averaged over the two sites) is required to pass Exercise 1. Where a participant's mark falls just below the 90% threshold, assessment of a third site may be permitted at the instructor's discretion. Record the mark for Exercise 1 in the space provided on the participant's Accreditation Assessment Results Summary Sheet (Appendix 1). Hand the field sampling sheets back to participants and work through any areas of difficulty with them. Participants may record their mark on the Module 2 assessment sheet.

#### Macroinvertebrate Sampling

#### **Exercise 2. Sample Collection**

#### Assessment

While completing Exercise 1, take each course participant aside and assess them collecting a riffle and edge sample as outlined in Exercise 2.

#### <u>Marking</u>

Award marks according to the marking scheme shown below. Participants receive full marks for each criterion correctly completed, and zero marks for those criteria incorrectly completed (i.e., a proportion of a mark cannot be given). A mark of 90% or greater (averaged over the two habitats) is required to pass Exercise 2. Record the marks for Exercise 2 in the space provided on the assessment sheet and on the Accreditation Assessment Results Summary Sheet. Hand the assessment sheet back to participants and work through any areas of difficulty with them.

As	sessable Criteria	Riffle Mark	Edge Mark
٠	10m of habitat sampled	/ 2	/ 2
٠	Available range of micro-habitats sampled	/ 5	/ 5
٠	Habitat sampled effectively	/ 3	/ 3
٠	Net rinsed when necessary	/ 2	/ 2
٠	Sample transferred successfully from net to sample container	/ 4	/ 4
٠	Sample container correctly labelled	/ 2	/ 2
٠	Sample preserved correctly with operator wearing protective gloves,		
	goggles and clothing (as required).	/ 2	/ 2
	Total	/ 20	/ 20

Note: A mark of 90% or greater is required in both exercises to pass Module 2.

## MODULE 3 – ASSESSMENT SHEET

#### Page 1 of 2

Name:

**Laboratory** 

Sample Processing

#### **Exercise 1. Laboratory Sub-Sample**

Collect a 200 organism sub-sample from both a riffle and edge sample following the South Australian laboratory sub-sampling protocols (see South Australian Sampling and Processing Manual for details). Retain the sub-sample residues in separate containers. Your instructor will sort through the sub-sample residues to ensure all the required animals have been removed. To pass you are required to recover an average of  $\geq$  95% of the total number of organisms from the two sub-samples.

(Exercise 1 Mark = %).

#### Macroinvertebrate Identification

#### **Exercise 2. Reference Collection Identification**

Identify macroinvertebrates in the reference collection to family taxonomic level, except for Oligochaeta (Class), Acarina (Order) and Chironomidae (Sub-family), using the keys recommended by Hawking (2000). You may also use the Interactive Guide to Australian Aquatic Invertebrates CD (Gunn *et al.* 1999) to aid in identifications.

Each taxon should be placed into a separate vial and the count for each recorded on a data sheet. To pass Exercise 2 you must achieve a "Percent Taxa Error" of  $\leq 5\%$ . A "Taxa Error" is defined as a mis-identification resulting in the loss or addition of a taxon. The "Percent Taxa Error" is the "Number of Taxa Errors" divided by the "Total Number of Original Taxa", multiplied by one hundred.

Percent Taxa Error		
Number of Taxa Errors (a)		
Total Number of Original Taxa (b)		
Percent Taxa Error ([a/b] x 100)		
Pass or Fail? (Pass if $\leq 5\%$ )		

(Exercise 2 Mark = %).

## MODULE 3 – ASSESSMENT SHEET

#### Page 2 of 2

#### **Exercise 3. Sample Identification**

Identify each of the macroinvertebrate sub-samples collected from the riffle and edge habitats to family taxonomic level, except for Oligochaeta (Class), Acarina (Order) and Chironomidae (Sub-family), using the keys recommended by Hawking (2000). You may also use the Interactive Guide to Australian Aquatic Invertebrates CD (Gunn *et al.* 1999) to aid in identifications.

Each taxon should be placed into a separate vial and the count for each recorded on a data sheet. Remember to record important sample information on the data sheet, such as the site number; river name; date; your name; habitat sampled, total number of organisms, total number of taxa etc.

To pass Exercise 3 you must achieve a "Percent Taxa Error" of  $\leq 5\%$  (averaged over the two samples).

(Exercise 3 Mark = %).

#### **Exercise 4. Scan**

Place the unsorted sample residues into separate trays and scan each for 15 minutes, collecting any taxa not found in the sub-samples. Macroinvertebrates collected during the scans should be kept separate from macroinvertebrates collected in the sub-samples and recorded on separate data sheets.

After you have scanned each of your samples for 15 minutes, your instructor will also scan the samples. To pass you are required to collect  $\ge$  90% of the taxa collected by your instructor (averaged over the two samples). Remember, only taxa not found in the original sub-sample should be collected.

(Exercise 4 Mark = %).

Note: To pass Module 3 you must pass Exercises 1 and 4 and achieve a "Percent Taxa Error" of  $\leq 5\%$  (averaged over Exercises 2 and 3).

## **MODULE 3 - INSTRUCTORS NOTES**

#### **Laboratory**

A list of materials and resources required for Module 3 is given below.

- Reference collection containing all families used in the creation of the South Australian AUSRIVAS models
- Riffle and edge macroinvertebrate sample
- Taxonomic keys for invertebrate taxa. See Key to Keys: A guide to keys and zoological information to identify invertebrates from Australian inland waters. Identification Guide No. 2, 2<sup>nd</sup> Edition.
  - (Hawking 2000). Visit the AUSRIVAS web site for a list of keys to families.
- Interactive Guide to Australian Aquatic Invertebrates, Compact Disk. 2nd Edition. (Gunn *et al.* 1999)
- Computer to run interactive identification CD.
- Invertebrate data sheets
- Staff with macroinvertebrate identification expertise
- Laboratory with stereo microscopes
- Forceps
- Vials
- Channel trays or grided dishes
- Counters
- Labels
- Sub-sampling box and vacuum pump
- Large white plastic sorting tray (for scan)
- Ethanol
- Squeeze bottles (for water and ethanol)
- Glycerol (dropper bottle)

#### Sample Processing

#### **Exercise 1. Laboratory Sub-Sample**

#### **Preparation**

Prepare laboratory with equipment required for sub-sampling (see above).

#### <u>Assessment</u>

Participants are required to collect a 200 organism sub-sample from both a riffle and edge sample following the South Australian laboratory sub-sampling protocols (See South Australian Sampling and Processing Manual for details). The South Australian protocol requires a 10% sub-sample with a minimum of 200 animals but 200 animals should be used for the training exercise so samples do not take excessively long to process. Ask participants to retain the sub-sample residues in separate containers. Sort through each participant's sub-sample residues and ensure all the required animals have been removed. Record the number of organisms missed by each

participant. Place these organisms into separate vials. These organisms should be added to the 200 organism sub-samples once Exercise 1 has been assessed.

#### <u>Marking</u>

To pass participants are required to recover an average of  $\ge 95\%$  of the total number of organisms from the two sub-samples. Record the mark for Exercise 1 in the space provided on the Accreditation Assessment Results Summary Sheet. Work through any areas of difficulty with participants.

#### Macroinvertebrate Identification

#### **Exercise 2. Reference Collection Identification**

#### **Preparation**

Prepare laboratory with equipment required for macroinvertebrate identification (see above). A reference collection containing all families used in the creation of the South Australian AUSRIVAS models is needed for this exercise.

#### Assessment

Participants are required to identify macroinvertebrates in the reference collection to family taxonomic level except for Oligochaeta (Class), Acarina (Order) and Chironomidae (Sub-family) using the keys recommended by Hawking (2000). The Interactive Guide to Australian Aquatic Invertebrates CD (Gunn *et al.* 1999) may also be used to aid in identifications. Once participants have completed the exercise, collect identification sheets, ensuring participant's names are written on the sheet.

#### <u>Marking</u>

To pass Exercise 2, participants must achieve a "Percent Taxa Error" of  $\leq 5\%$ . A "Taxa Error" is defined as a mis-identification resulting in the loss or addition of a taxon. The "Percent Taxa Error" is the "Number of Taxa Errors" divided by the "Total Number of Original Taxa", multiplied by one hundred.

Percent Taxa Error			
Number of Taxa Errors (a)			
Total Number of Original Taxa (b)			
Percent Taxa Error ([a/b] x 100)			
Pass or Fail? (Pass if $\leq 5\%$ )			

Record the mark for Exercise 2 in the space provided on the Accreditation Assessment Results Summary Sheet. Hand sheets back to participants and work through with them any areas of difficulty.

#### **Exercise 3. Sample Identification**

#### **Preparation**

Prepare laboratory with equipment required for macroinvertebrate identification (see above). Two samples are required for this exercise, one from a riffle habitat and another from a stream edge habitat.

#### <u>Assessment</u>

Participants are required to identify macroinvertebrate sub-samples collected from both the riffle and edge habitats to family taxonomic level, except for Oligochaeta (Class), Acarina (Order) and Chironomidae (Sub-family) using the keys recommended by Hawking (2000). The Interactive Guide to Australian Aquatic Invertebrates CD (Gunn *et al.* 1999) may also be used to aid in identifications. Once participants have completed the exercise, collect identification sheets, ensuring participant's names are written on the sheet.

#### <u>Marking</u>

To pass Exercise 3, participants are required to achieve a "Percent Taxa Error" of  $\leq 5\%$  (averaged over the two samples). Record the mark for Exercise 3 in the space provided on the Accreditation Assessment Results Summary Sheet. Hand sheets back to participants and work through with them any areas of difficulty.

#### **Exercise 4. Scan**

#### **Preparation**

Prepare laboratory with equipment required for a scan (see above).

#### <u>Assessment</u>

Participants are required to place their unsorted sample residues into separate trays and scan each for 15 minutes, collecting any taxa not found in the sub-samples. Macroinvertebrates collected during the scans should be kept separate from macroinvertebrates collected in the sub-samples and recorded on separate data sheets.

After the participants have scanned each of the samples for 15 minutes, re-scan the samples for another 15 minutes, collecting taxa not found in the original sub-samples.

#### <u>Marking</u>

To pass participants are required to collect  $\ge 90\%$  of the taxa collected by the instructor (averaged over the two samples). Record the mark for Exercise 4 in the space provided on the Accreditation Assessment Results Summary Sheet. Show participants any taxa missed in their scan.

Note: To pass Module 3 participants must pass Exercises 1 and 4 and achieve a "Percent Taxa Error" of  $\leq 5\%$  (averaged over Exercises 2 and 3).

## MODULE 4 – ASSESSMENT SHEET

#### Page 1 of 4

Name:

#### **AUSRIVAS Predictive Models**

Using the Predictive Models

#### **Exercise 1. Single-Season Predictive Model**

a) Determine the habitat predictor variables required by the \_\_\_\_\_

\_\_\_\_\_ AUSRIVAS model and list below.

Predictor Variables Used:

- b) Place macroinvertebrate and habitat data into the format required by AUSRIVAS.
- c) Import the bug and habitat files into AUSRIVAS and run model. Save the file on the floppy disk provided. Call the file "your name\_single\_model.AUS".
- d) Export the AUSRIVAS sheet labelled Predicted/Collected containing the Observed to Expected (O/E) ratios and open in Excel.
- e) Save this file as an Excel spreadsheet on the floppy disk provided. Call the file "your name\_single\_oe.xls". Ensure your name is written on the disk.

A mark of 90% or greater is required to pass Exercise 1.

(Exercise 1 Mark = %).

## MODULE 4 – ASSESSMENT SHEET

#### Page 2 of 4

#### **Exercise 2. Combined-Season Predictive Model**

Predictor Variables Used:

- b) Combine the macroinvertebrate data for autumn and spring.
- c) Combine the habitat data for autumn and spring.
- d) Place macroinvertebrate and habitat data into the format required by AUSRIVAS.
- e) Import the bug and habitat files into AUSRIVAS and run model. Save the file on the floppy disk provided. Call the file "your name\_combined\_model.AUS".
- f) Export the AUSRIVAS sheet labelled Predicted/Collected containing the Observed to Expected (O/E) ratios and open in Excel.
- g) Save this file as an Excel spreadsheet on the floppy disk provided. Call the file "your name\_combined\_oe.xls". Ensure your name is written on the disk.

A mark of 90% or greater is required to pass Exercise 2.

(Exercise 2 Mark = %).

## **MODULE 4 – ASSESSMENT SHEET**

#### Page 3 of 4

#### Interpreting the Results

#### **Exercise 3. Site Assessment**

Using the information provided, complete the summary table below.

Site	Overall Site Assessment (Band)	Habitat used for Overall Site Assessment	Taxa Predicted to occur but not collected in the habitat used for overall site assessment

Note: If both riffle and edge habitats are placed into the same band, refer to the habitat with the lowest O/E taxa ratio.

One mark is awarded for each section of the table completed correctly. A mark of 90% or greater is required to pass Exercise 3.

(Exercise 3 Mark = \_\_\_%).

## **MODULE 4 – ASSESSMENT SHEET**

#### Page 4 of 4

#### **Exercise 4. Interpreting Results**

List 5 of the steps that one can take when interpreting AUSRIVAS results.

1.			
2.			
3.			
4.			
5.			

One mark is awarded for each step listed. A mark of 4 out of 5 (80%) or greater is required to pass Exercise 4.

(Exercise 4 Mark = \_\_\_%).

Note: To pass Module 4 you must pass all four exercises.

## **MODULE 4 - INSTRUCTORS NOTES**

#### AUSRIVAS Predictive Models

A list of materials and resources required for Module 4 is given below.

- Predictive Model Manual hard copy
- Macroinvertebrate and habitat data for two sites assessed as mildly impacted
- AUSRIVAS combined season riffle and edge model outputs for five sites
- Access to AUSRIVAS models (password)
- Computers with AUSRIVAS software
- Access to the Internet
- Floppy disk for each participant containing the above data

Note: The AUSRIVAS Training and Accreditation CD contains demonstration data from the ACT that may prove useful when developing data sets for South Australia.

#### Using the Predictive Models

#### **Exercise 1. Single-Season Predictive Model**

#### <u>Preparation</u>

Prepare a data set comprising macroinvertebrate and habitat data collected from two mildly impacted test sites. The macroinvertebrate data may be from any habitat for which there is a single season model and should be saved as an Excel spreadsheet. Photocopies of the field sampling sheets can be used to provide the habitat data.

#### <u>Assessment</u>

Participants are required to manipulate the data into the format required by AUSRIVAS and run the data through a single season model.

#### <u>Marking</u>

Award marks for each question according to the marking scheme shown below. Participants receive full marks for each criterion correctly completed, and zero marks for those criteria incorrectly completed (i.e., a proportion of a mark cannot be given). A mark of 90% or greater is required to pass Exercise 1. Record the mark for Exercise 1 in the space provided on the Accreditation Assessment Results Summary Sheet. Work through any areas of difficulty with participants.

Assessable Criteria	Marks	Comments
Predictor Variables Correct	/ 5	
Macroinvertebrate data in correct format:		
- Adults and larvae combined	/ 2	
- Bug codes correct	/ 5	
- Zeros added and no blanks found	/ 2	
- Data with sites as rows and bugs as columns	/ 2	
- File saved as text (tab delimited) file	/ 1	
Habitat data in correct format:		
- Habitat codes correct	/ 3	
- Zeros added and no blanks found	/ 2	
- Data with sites as rows and habitat variables as columns	/ 2	
- Sites in the same order as the bug data	/ 2	
- File saved as text (tab delimited) file	/ 1	
• Imported the bug file into AUSRIVAS successfully	/ 2	
• Imported the habitat file into AUSRIVAS successfully	/ 2	
• Choose the correct AUSRIVAS model i.e. region, season and	/ 3	
habitat		
• Exported the AUSRIVAS predicted/collected sheet successfully	/ 2	
Opened the AUSRIVAS predicted/collected sheet in excel and		
saved the file as an excel spreadsheet	/ 2	
Total	/ 38	

#### **Exercise 2. Combined-Season Predictive Model**

#### **Preparation**

Prepare a data set comprising macroinvertebrate and habitat data collected from two mildly impacted test sites. The macroinvertebrate data should be from the same habitat and include both autumn and spring data. The data should be saved as an Excel spreadsheet. Photocopies of the autumn and spring field sampling sheets can be used to provide the habitat data.

#### <u>Assessment</u>

Participants are required to manipulate the data into the format required by AUSRIVAS and run the data through a combined season model.

#### <u>Marking</u>

Award marks for each question according to the marking scheme shown below. Participants receive full marks for each criterion correctly completed, and zero marks for those criteria incorrectly completed (i.e., a proportion of a mark cannot be given). A mark of 90% or greater is required to pass Exercise 2. Record the mark for Exercise 2 in the space provided on the Accreditation Assessment Results Summary Sheet. Work through any areas of difficulty with participants.

Assessable Criteria	Marks	Comments
Predictor Variables Correct	/ 5	
• Macroinvertebrate data in correct format:		
- Autumn and spring macroinvertebrate data combined	/ 5	
- Adults and larvae combined	/ 2	
- Bug codes correct	/ 5	
- Zeros added and no blanks found	/ 2	
- Data with sites as rows and bugs as columns	/ 2	
- File saved as text (tab delimited) file	/ 1	
• Habitat data in correct format:		
- Autumn and spring habitat data combined	/ 5	
- Habitat codes correct	/ 3	
- Zeros added and no blanks found	/ 2	
- Data with sites as rows and habitat variables as columns	/ 2	
- Sites in the same order as the bug data	/ 2	
- File saved as text (tab delimited) file	/ 1	
• Imported the bug file into AUSRIVAS successfully	/ 2	
• Imported the habitat file into AUSRIVAS successfully	/ 2	
• Choose the correct AUSRIVAS model i.e. region, season and	/ 3	
	1.2	
• Exported the AUSRIVAS predicted/collected sheet successfully	/ Z	
• Opened the AUSRIVAS predicted/collected sheet in excel and	12	
saved the file as an excel spreadsneet	/ 2	
	/ 48	

#### Interpreting the Results

#### **Exercise 3. Site Assessment**

#### **Preparation**

Prepare two data sets comprising AUSRIVAS combined season riffle and edge model outputs for five sites. As the first part of this exercise is designed to test whether course participants know the rules for combining bands, sites exhibiting different band assessments between habitats should be used.

#### <u>Assessment</u>

Participants are required to complete the summary table, indicating the:

• Overall site assessment - determined by combining assessments from the riffle and edge habitats.

The rules for combining assessments from different habitats are as follows. Where the bandings from both habitats allocate the site to the same band, then that is the final band allocation for the site. Where there is a mismatch in the band allocation from the two habitats, then allocate the site to the band that is farther from band A. In the rare event that the alternative bands are band B and band X, allocate to band B, because this is the most precautionary approach.

- Habitat used for the overall site assessment. If both riffle and edge habitats are placed into the same band, the habitat with the lowest O/E taxa ratio should be referred to.
- Taxa predicted to occur but not collected in the habitat used for the overall site assessment.

#### <u>Marking</u>

Award marks for each question according to the marking scheme shown below. Participants receive full marks for each criterion correctly completed, and zero marks for those criteria incorrectly completed (i.e., a proportion of a mark cannot be given). A mark of 90% or greater is required to pass Exercise 3. Record the mark for Exercise 3 in the space provided on the Accreditation Assessment Results Summary Sheet. Work through any areas of difficulty with participants.

As	sessable Criteria	Marks (1 mark per site)	Comments
•	Overall site assessment correct	/ 5	
٠	Habitat used in overall site assessment correct	/ 5	
٠	Taxa predicted but not collected correct	/ 5	
	Total	/ 15	

#### **Exercise 4. Interpreting Results**

#### <u>Assessment</u>

Participants are required to list 5 of the steps that one can take when interpreting AUSRIVAS results. Steps include:

- Check for data entry errors bug data and habitat data.
- Check "Unused Bugs" for any bugs which should have been used by AUSRIVAS but were maybe given the wrong code.
- Examine O/E values in relation to the band width e.g., if a site is assigned Band B, how far below reference is the site?
- Band X may require further examination possible biological "hot spot", nutrient enrichment, constant flow in normally intermittent stream etc.
- Examine the list of taxa predicted to occur but not collected.
- Examine field data sheets for impact indicators, including
  - poor water quality
  - poor habitat
  - land use impacts
  - point source pollution
  - non-point source pollution
  - dams/barriers upstream
  - recent flooding or low flows
  - under-sampling
  - poor live-sorting conditions
- May need to draw a conclusion of "no reliable assessment possible"

Steps other than those listed above may be deemed acceptable.

#### <u>Marking</u>

Award 1 mark for each of the 5 steps listed. A mark of 4 out of 5 (80%) or greater is required to pass Exercise 4. Record the mark for Exercise 4 in the space provided on the Accreditation Assessment Results Summary Sheet. Work through any areas of difficulty with participants.

Note: To pass Module 4 participants must pass all four exercises.

## **REFERENCES**

Gunn, B., Cranston, P.S., Dimitriadis, S. & Trueman, J.W.H. (1999). *Interactive guide to Australian Aquatic Invertebrates*. Windows edn 2. CSIRO, Land & Water Resources Research & Development Corporation, & Environment Australia: Canberra.

Hawking J.H. (2000) *Key to Keys: A Guide to keys and zoological information to identify invertebrates from Australian inland waters, Identification Guide No. 2*, 2nd Edn. Cooperative Research Centre for Freshwater Ecology, Albury, NSW.

## **APPENDICES**

## Appendix 1 Accreditation Assessment Results Summary Sheet

Participants Name:

MODULE	<b>MARK (%)</b>	MARK (%) REQUIRED	PASS/FAIL	COMMENTS
		TO PASS		
MODULE 1				
Exercise 1		≥90		
MODULE 2				
Exercise 1		≥90		
Exercise 2		≥90		
MODULE 3				
Exercise 1		≥95		
Exercise 2		≥95		
Exercise 3		≥95		
Exercise 4		≥90		
MODULE 4				
Exercise 1		≥90		
Exercise 2		≥90		
Exercise 3		≥90		
Exercise 4		≥80		

MODULE	PASS/FAIL
Module 1	
Module 2	
Module 3	
Module 4	

Organisation Conducting Course:

Instructor's Name:

Instructor's Signature:

Date: